Microbiological and Immunological Studies on Brucellosis in a Hospital in Al-Madinah Al-Munawarah

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Abstract

**Purpose:** The study was conducted to estimate the prevalence of brucellosis in Prince Sultan Armed Forces Hospital at Al-Madinah Al-Munawarah, Saudi Arabia. The aim was also to determine the most prevalent species of Brucella and to make a comparison between culture and serological methods in diagnosis and to evaluate the levels of sIL-2R and/or IFN-γ production to be used as markers of treatment efficacy.

**Study Design:** Cross-sectional Study

**Subjects and Methods:** This study was conducted on 65 patients with male: female ratio (2:1) suspected of having brucellosis. It was carried out using slide agglutination test for detection of anti-Brucella antibodies. Also, we estimated anti-Brucella IgG and IgM antibody levels in the sera of examined patients using ELISA. Quantization of human IFN-γ was performed.

**Results:** The total incidence of brucellosis was 92.3%. The incidence among males (95.2%) was higher than that of female (87%). Brucellosis was detected in all age groups. Most of brucellosis patients were recovered during the period from January to June. Consumption of milk products, heating raw milk and milking animals were the highest risks with an incidence of 100% followed by drinking raw milk with an incidence of 95% while cutting raw meat and animal contact were the less risk with an incidence of 80% and 67%, respectively. The most prevalent species among examined patients was B. melitensis (86%) and B. abortus (6%). Brucellosis patients had 63% and 83% of anti Brucella IgG and IgM, respectively. The highest (%) of patients having positive IgG and IgM levels in their sera were among 1/160 standard tube agglutination test (SAT) antibody titer group brucellosis patients were having positive levels of IFN-γ. All of them belonged to 1/80 antibody titer group. The mean IFN-γ levels according to SAT antibody titer were 224.25, 102 and 69.3 pg/ml, respectively.

**Conclusion:** Eradication of human brucellosis depends on the eradication of animal brucellosis. In countries like the Kingdom of Saudi Arabia, where brucellosis is endemic; rapid, sensitive and highly specific diagnostic methods are required to make early diagnosis and prevent resistance as there is an overlap in therapy.

1. Introduction

Brucellosis is a major zoonotic disease that poses a serious public health and economic problem worldwide (Corbel, 1997). The heaviest disease burden lies in countries of the Mediterranean basin and Arabian area. The disease also remains endemic in many countries, particularly in India, Mexico, and South and Central America; however the disease

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incidence and prevalence rates vary widely among nations (Khan et al., 2001). It is estimated that the annual incidence of brucellosis in Saudi Arabia is 21.4/100,000 population (Pappas et al., 2006). The disease is caused by small Gram-negative bacteria belonging to the genus *Brucella*. It is non-motile, non-encapsulated cocco-bacilli, which functions as facultative intracellular parasite. There are a few different species of *Brucella*, each with slightly different host specificity (Meyer, 1990). *B. melitensis* that is found primarily in goats, sheep and camels and *B. abortus* that is found primarily in cattle are the principal cause of human brucellosis (Plommet & Verger, 1996), however *B. Melitensis*, the most invasive, produces the most severe disease (Al-Eissa, 1999). The disease is transmitted from animals to humans by direct contact with infected animals, consumption of their infected dairy products, or inhalation of aerosols (Kurdoglu et al., 2010). Other than the possibility of transmission through blood transfusion and organ or bone marrow transplantation, brucellosis is not readily transmitted between human beings (Khan et al., 2001). Brucellosis may be seen in any age group, but it still involves young and middle aged adults more frequently with predominance among males.

The majority of brucellosis cases were attributed to direct contact with animals or their products (Hizel et al., 2007). Although it is thought that brucellosis causes fewer spontaneous abortion in humans than it does in animals due to the lack of erythritol, a preferential medium and growth factor for *Brucella*, in the human placenta and the presence of anti-Brucella activity in human amniotic fluid, there is some evidence that it may cause a higher rate of complications, such as abortion, preterm labor, and intrauterine fetal demise, more frequently than do other bacterial infections (Hizel et al., 2007). In humans, brucellosis behaves as a systemic infection with a very heterogeneous clinical spectrum. The disease usually presents as fever with no apparent focus, although there are focal forms in 20–40% of cases. As the clinical picture of human brucellosis is fairly non-specific, a definitive diagnosis requires isolation of the causative organism or the demonstration of high levels of specific antibodies, or seroconversion (Ruiz-Mesa et al., 2005).

During the first week of infection, IgM antibodies against lipopolysaccharide antigens appear in the serum, followed by IgG antibodies as early as the second week. Both antibody isotypes peak during the fourth week, and the use of antibiotics was associated with a decline of both IgM and IgG class antibodies. The diagnosis of brucellosis is based on a detailed history obtained from the patients and the isolation of *Brucella* spp. from blood cultures are accepted as the standard method. The definitive diagnosis of brucellosis requires isolation of the bacterium from blood, bone marrow or tissue samples. Although the sensitivity of blood culture method varies depending on the individual laboratory practices, quantity of bacteria in blood and the methods used, it changes between 15-70 percent. Bone marrow culture is considered as the gold standard for the diagnosis of brucellosis, since the relatively high concentration of *Brucella* in reticuloendothelial system enables the detection of the organism. The standard tube agglutination test (SAT) is the most widely used serologic test for the confirmation of human brucellosis. Otherwise, enzyme immunoassays (ELISA) which measure specific IgM, IgG and IgA antibodies, allow for a better interpretation of the clinical situation and overcome the false negativit/positivities which may arise in SAT (Alişka, 2008). The whole-blood interferon-gamma (IFN-γ) assay is a quantitative in vitro assay for a
direct read-out of Ag-specific cell-mediated immune (CMI) responses to infectious diseases. The IFN-γ assay is robust in severe intracellular infections like Brucella (Riber et al., 2011). For all the above reasons and as human brucellosis has a serious medical impact in Saudi Arabia, the study had been conducted to estimate the prevalence of brucellosis in a Saudi hospital in Al-Madinah Al-Munawarah, determine the most prevalent species of Brucella (Brucellameliitensis, Brucella abortus), compare the culture and serological methods in diagnosis of those patients with signs and symptoms of brucellosis, and to evaluate levels of sIL-2R and/or IFN-γ production during infection with species of Brucella and to detect (if possible) their possibility to be used as markers of treatment efficacy in individuals suffering from brucellosis.

The Kingdom of Saudi Arabia imports millions of sheep and goats each year for the Pilgrimage season. Thus, the eradication of human brucellosis depends on the eradication of animal brucellosis, and the formation of a national program for controlling brucellosis. Prevention of brucellosis can be best achieved by minimizing human contact with infected animals or their products. Hand washing, eye protection, protective clothing, tool/surface disinfection, infected animal slaughter and destruction are some of the more important measures to be taken in farms and other premises. Raw milk and meat consumption should be avoided to prevent infection of the general population (Corbel, 1997). In addition to that, in countries like the Kingdom of Saudi Arabia, where brucellosis is endemic; rapid, sensitive and highly specific diagnostic methods are required to make early diagnosis and prevent resistance as there is an overlap in therapy (Al-Eissa, 1999).

2. Methodology and Procedures

Patients

This study was conducted on a total of 65 patients (42 Male & 23 Female) suspected of having brucellosis. All the patients were admitted to Prince Sultan Armed Forces Hospital at Al-Madinah Al-Munawrah during the period of November 2015 to June 2016. This study included 18 control subjects (10 Male & 8 Female) with apparently no evidence of infection with brucellosis (based on absence of brucellosis symptoms currently and no previous history of brucellosis) as negative controls. 5ml of blood was obtained from patients and controls. The collected serum was stored at -20°C for further tests.

Determination of Brucella Species by Febrile Antigen Kit

All sera of patients and controls were tested for Brucella antibody (AB) using febrile antigen kit from Plasmatec Laboratory products, UK. Stained antigen suspensions (Brucella abortus 5 ml, Brucellameliitensis 5 ml, Positive control 0.5 ml and Negative control 0.5 ml) are for the identification and quantitative determination of specific antibodies in human sera following infection with certain Brucella pathogens. Febrile antigens were suitable for slide agglutination test against human sera for detection of these agglutinins. Stained antigen suspension were killed bacteria, stained to enhance the reading of agglutination tests. If sufficient homologous antibodies were present, they would cause the suspension to visibly agglutinate.
Rapid Slid Titration

It was performed according to the manufacturer’s directions. 0.08 ml, 0.04 ml, 0.02 ml, 0.01 ml and 0.005 ml of undiluted serum were dispensed onto a row of 3 cm diameter circles by a micropipette. If Agglutination reactions are observed, the titer was calculated according to the following approximate values: 0.08 ml = 1:20, 0.04 ml = 1:40, 0.02 ml = 1:80, 0.01 ml = 1:160, 0.005 ml = 1:320

Detection of Brucella IgM / IgG by Novagnost ELISA Kit

All sera of patients and controls were tested for Brucella IgM AB and Brucella IgG AB using novagnost kit from Nova Tec, Germany.

Photo (1): Detection of anti-Brucella IgG antibodies in examined patients using ELISA

Photo (2): Detection of anti-Brucella IgM antibodies in examined patients using ELISA

Immunological Assay Human IFN-γ:

The kit used for the quantitation of IFN-γ was provided by Quantikine (R & D systems, Inc., Minneapolis, USA). This assay employs the quantitative sandwich enzyme immunoassay technique. It was performed manually according to the manufacturer's directions as shown in table (1).
Statistical Analysis

All data were analyzed using GRAPHPAD PRISM software (GraphPad, San Diego, CA). Data were analyzed using ANOVA (multiple groups), and multiple comparisons between the groups were performed using Newman–Keuls method after ANOVA. P < 0.05 was considered as statistically significant for all experiments. All values were presented as the mean ± SD.

3. Results and Discussion

Effect of Gender on Brucellosis:

As shown in figure (2) the slide agglutination test revealed that 60 patients were brucellosis positive with a total incidence of 92.3%. Out of 42 male patients, 40 brucellosis cases were recovered with an incidence of 95.2%. On the other hand, 20 brucellosis cases were identified from 23 female patients with an incidence of 87%. It was of interest to note that male: female ratio in this study was 2:1 “P-value 0.231”
Effect of Age on Brucellosis

The results recorded in table (2) show that out of 65 examined patients, 3 patients were belonging to age (10-20 years old), 9 patients were belonging to age (20-30 years old), 14 patients were belonging to age (30-40 years old), 25 patients were belonging to age (40-50 years old) and 14 patients were belonging to age (50-60 years old). Based on results of slide agglutination test, the incidence of brucellosis among these patients groups were 100%, 89%, 93%, 88% and 100%, respectively.

Table (2): Distribution of brucellosis cases by age of patients

<table>
<thead>
<tr>
<th>Age</th>
<th>Total No. of examined patients</th>
<th>No. (%) Of Positive</th>
<th>Mean±S.D</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>10-20</td>
<td>3</td>
<td>3 (100%)</td>
<td>19.3±0.577</td>
<td>0.695</td>
</tr>
<tr>
<td>20-30</td>
<td>9</td>
<td>8 (89%)</td>
<td>27.4±2.26</td>
<td>0.695</td>
</tr>
<tr>
<td>30-40</td>
<td>14</td>
<td>13 (93%)</td>
<td>36.8±2.35</td>
<td>0.695</td>
</tr>
<tr>
<td>40-50</td>
<td>25</td>
<td>22 (88%)</td>
<td>44.9±3.05</td>
<td>0.695</td>
</tr>
<tr>
<td>50-60</td>
<td>14</td>
<td>14 (100%)</td>
<td>56.9±3</td>
<td>0.695</td>
</tr>
<tr>
<td>Total</td>
<td>65</td>
<td>60 (92.3%)</td>
<td>42.3±11.11</td>
<td></td>
</tr>
</tbody>
</table>

Source: Authors

Distribution of Brucellosis Cases by Season

From Figure (3), it is noticed that the most of brucellosis patients were recovered during the period from January to June with an incidence of 100% for all months except March was 83%. On the other hand, the examined patients during November and December were negative for brucellosis using slide agglutination test "P-value 0.000".

Figure (3): Distribution of brucellosis cases by season
Effect of Risk Factors on Brucellosis

It can be concluded from figure (4) that there were different risk factors for brucellosis. Consumption of milk products, heating raw milk and milking animals were the most risk with an incidence of 100% followed by drinking raw milk with an incidence of 95%. Meanwhile, cutting raw meat and animal contact were the less risk with an incidence of 80% and 67%, respectively "P-value 0.142".

Figure (4): Distribution of brucellosis cases by risk factor

Titers of Anti-Brucella Antibodies in Brucellosis Patients

Anti-Brucella antibodies of brucellosis cases were evaluated also using rapid slid agglutination test. From data available in table (3), it is evident that anti-Brucella antibodies titers reach to 80 in fourteen cases, 160 in thirty four cases and 320 in twelve cases.

<table>
<thead>
<tr>
<th>SAT anti-Brucella antibody titers</th>
<th>No. (%) of Positive cases</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>1/80</td>
<td>14 (22%)</td>
<td></td>
</tr>
<tr>
<td>1/160</td>
<td>34 (52%)</td>
<td></td>
</tr>
<tr>
<td>1/320</td>
<td>12 (18.5%)</td>
<td>0.000</td>
</tr>
<tr>
<td>1/640</td>
<td>0 (0%)</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>60 (92.3%)</td>
<td></td>
</tr>
</tbody>
</table>

Source: Authors

Results Of Brucella Species Detection

Using slide latex agglutination test are presented in figure (5). They illustrated that the most prevalent species among examined patients was B. melitensis (86%) with P-value 0.551. On the other hand, B. abortus was detected from only (6%) of examined cases with P-value 0.000.
Results of Detection of Anti-Brucella IgM Antibodies and anti- Brucella IgG Antibodies in Examined Patients Using (ELISA)

Estimation of Brucella specific IgG and IgM antibody levels in the sera of examined patients as shown in table (4 & 5) and photo (1 & 2) revealed that the sera of 38 (63%) brucellosis patients were containing anti- Brucella Ig G antibodies and 22 (37%) were negative. However, the sera of 4 (80%) patients free from brucellosis were containing anti-Brucella IgG antibodies and only one (20%) was negative. On the other hand, the sera of 50 (83%) brucellosis patients were containing anti- Brucella Ig M antibodies and 10 (17%) of them were negative. However, the sera of 2 (40%) patients free from brucellosis were containing anti- Brucella IgM antibodies and 3 (60%) of them were negative. The data presented in table (6) recorded 34 (57%) patients with brucellosis and only one (20%) patient free from brucellosis were containing IgG/IgM combined antibodies in their sera. It is of interest to note that 6 (10%) patients with brucellosis were negative for both.

Table (4): Results of detection of anti- Brucella IgG antibodies in examined patients using ELISA comparing with SAT

<table>
<thead>
<tr>
<th>Patients</th>
<th>IgG No. (%)</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patients with brucellosis</td>
<td>38 (63.3%)</td>
<td>22 (36.7%)</td>
<td>63.3%</td>
<td>0.454</td>
</tr>
<tr>
<td>(60)2</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Patients free from</td>
<td>4 (80%)</td>
<td>1 (20%)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>brucellosis (5)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Source: Authors
Table (5): Results of detection of anti-Brucella IgM antibodies in examined patients using ELISA comparing to SAT

<table>
<thead>
<tr>
<th>Patients</th>
<th>IgG/M Combined No. (%)</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patients with Brucellosis having IgG/M combined positive or negative (40)</td>
<td>34 (85%) 6 (15%)</td>
<td>85%</td>
<td>0%</td>
<td>0.675</td>
</tr>
<tr>
<td>Patients free from brucellosis having IgG/M combined positive or negative (1)</td>
<td>1 (100%) 0 (0%)</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Source: Authors

Table (6): Results of detection of anti-Brucella IgG/M combined Antibodies in examined patients using ELISA comparing to SAT

<table>
<thead>
<tr>
<th>Patients</th>
<th>IgM No. (%)</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patients with Brucellosis (60)</td>
<td>50 (83.3%) 10 (16.7%)</td>
<td>83.3%</td>
<td>60%</td>
<td>0.020</td>
</tr>
<tr>
<td>Patients free from brucellosis (5)</td>
<td>2 (40%) 3 (60%)</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Source: Results of quantification of human IFN-γ:

Interpretation of the readings is based on the tested serum sample was considered as positive if its ELISA absorbance value exceeds the mean value of the negative control samples by more than two standard deviations (Gast et al., 1997). By comparing ELISA absorbance values of IFN-γ in sera of examined patients with the calculated cut off value (0.261) we resulted in among patients with brucellosis, 9 cases were having positive levels of IFN-γ while all healthy humans were having negative levels of IFN-γ (table 7).
Table (7): Evaluation of IFN-γ levels of examined patients based on cut off value

<table>
<thead>
<tr>
<th>IFN-γ levels</th>
<th>No. (%) of patients</th>
<th>Range</th>
<th>Mean±S.D</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive IFN-γ</td>
<td>9 (14%)</td>
<td>0.21</td>
<td>0.345±0.08</td>
<td></td>
</tr>
<tr>
<td>Levels</td>
<td></td>
<td></td>
<td></td>
<td>0.000</td>
</tr>
<tr>
<td>Negative IFN-γ</td>
<td>56 (86%)</td>
<td>0.26</td>
<td>0.134±0.04</td>
<td></td>
</tr>
<tr>
<td>Levels</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>65</td>
<td>0.48</td>
<td>0.164±0.09</td>
<td></td>
</tr>
</tbody>
</table>

Source: Authors

Sero-Prevalence of Brucellosis in Humans

*Relationship between the results of SAT and ELISA (IgM, IgG and IFN-γ) in detection of anti-Brucella antibodies:* Results of SAT as well as ELISA (IgG, IgM and IFN-γ) were recorded and tabulated in table (8). 95%, 83%, 71% and 12% respectively were brucellosis positive males based on these tests. In addition to 87%, 74%, 52% and 17% respectively were brucellosis positive females based on these tests (figure 6).

Table (8): Relationship between the results of SAT and ELISA (IgM, IgG and IFN-γ) in detection of anti-Brucella antibodies (the gold standard method for detection of brucellosis is SAT).

<table>
<thead>
<tr>
<th>Gender</th>
<th>No. of examined patients</th>
<th>SAT</th>
<th>ELISA</th>
<th>IFN-γ</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>+ve</td>
<td>IgM</td>
<td>+ve</td>
</tr>
<tr>
<td>Male</td>
<td>42</td>
<td>40</td>
<td>2</td>
<td>35</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(95%)</td>
<td>(5%)</td>
<td>(83%)</td>
</tr>
<tr>
<td>Female</td>
<td>23</td>
<td>20</td>
<td>3</td>
<td>17</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(87%)</td>
<td>(13%)</td>
<td>(74%)</td>
</tr>
<tr>
<td>Total</td>
<td>65</td>
<td>60</td>
<td>5</td>
<td>52</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(92%)</td>
<td>(8%)</td>
<td>(80%)</td>
</tr>
</tbody>
</table>

Source: Authors

Figure (6): Relationship between the results of SAT and ELISA (IgM, IgG and IFN-γ) in detection of anti-Brucella antibodies
Graphical Comparison of SAT with ELISA for Detection of anti-Brucella Antibodies in Sera

As indicated, 40 male patients with brucellosis based on SAT were examined using ELISA for the presence of anti-Brucella IgG and IgM antibodies and IFN-γ in their sera. The results revealed that 85% of them were having positive IgM titers, 73% were having positive IgG titers and 12.5% were having positive IFN-γ levels. However, 80%, 45% and 20% of female patients with brucellosis were having positive levels of IgM, IgG and IFN-γ, respectively (table 9 and figure 7). From the same table, it is noticed that B. abortus was detected in three male patients and one female patient with brucellosis. The rest of brucellosis patients were infected with B. melitensis. The sensitivity, specificity, negative predictive values and positive predictive values of ELISA tests used in this study are shown in table (10).

Table (9): Distribution of Brucella antibody findings among 65 patients tested by different methods

<table>
<thead>
<tr>
<th>Patients groups</th>
<th>No. (%) of Positive SAT</th>
<th>Positive ELISA</th>
<th>Brucella species</th>
<th>IFN-γ</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>IgM</td>
<td>IgG</td>
<td>B.melitensis</td>
</tr>
<tr>
<td>Male brucellosis patients</td>
<td>40</td>
<td>34</td>
<td>29</td>
<td>37</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(85%)</td>
<td>(73%)</td>
<td>(92.5%)</td>
</tr>
<tr>
<td>Female brucellosis patients</td>
<td>20</td>
<td>16</td>
<td>9</td>
<td>19</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(80%)</td>
<td>(45%)</td>
<td>(95%)</td>
</tr>
<tr>
<td>Male free from brucellosis</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Female free from brucellosis</td>
<td>0</td>
<td>1</td>
<td>3</td>
<td>0</td>
</tr>
</tbody>
</table>

Source: Authors

Figure (7): Distribution of Brucella antibody findings among 65 patients tested by different methods
Table (10): Sensitivity, specificity, negative predictive values and positive predictive values of ELISA used in this study comparing to SAT

<table>
<thead>
<tr>
<th>ELISA</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>PPV *</th>
<th>NPV**</th>
</tr>
</thead>
<tbody>
<tr>
<td>IgG</td>
<td>63.3%</td>
<td>20%</td>
<td>90.5%</td>
<td>4.4%</td>
</tr>
<tr>
<td>IgM</td>
<td>83.3%</td>
<td>60%</td>
<td>96.2%</td>
<td>23.1%</td>
</tr>
<tr>
<td>IFN-γ</td>
<td>15%</td>
<td>100%</td>
<td>100%</td>
<td>8.9%</td>
</tr>
</tbody>
</table>

Source: Authors

* PPV means positive predictive value
** NPV means negative predictive value

Distribution of Positive Results of ELISA According to SAT Titers

It can be concluded from table (11) that there were differences in ELISA results between 1/80, 1/160 and 1/320 antibody titer groups. The highest (%) of patients having positive IgG and IgM levels in their sera were in 1/160 antibody titer group. It was also noticed that all positive IFN-γ levels were belonging to 1/80 antibody titer group. The mean IFN-γ levels according to SAT antibody titers were 224.25, 102 and 69.3 pg / ml, respectively (table 12).

Table (11): Distribution of positive results of ELISA according to SAT titers

<table>
<thead>
<tr>
<th>ELISA</th>
<th>1/80 (No. (%) of Positive)</th>
<th>1/160 (No. (%) of Positive)</th>
<th>1/320 (No. (%) of Positive)</th>
<th>Total (No. (%) of Positive)</th>
</tr>
</thead>
<tbody>
<tr>
<td>IFN-γ</td>
<td>9 (64%)</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
<td>9 (15%)</td>
</tr>
<tr>
<td>Ig M</td>
<td>12 (86%)</td>
<td>30 (88%)</td>
<td>8 (67%)</td>
<td>50 (83%)</td>
</tr>
<tr>
<td>Ig G</td>
<td>9 (64%)</td>
<td>23 (68%)</td>
<td>6 (50%)</td>
<td>38 (63%)</td>
</tr>
</tbody>
</table>

Source: Authors

Table (12): The mean IFN-γ levels (pg/ml) in relation to the SAT antibody titers

<table>
<thead>
<tr>
<th>Mean ELISA absorbance value</th>
<th>SAT titers</th>
<th>Mean IFN-γ levels (pg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.141</td>
<td>Control</td>
<td>105.75</td>
</tr>
<tr>
<td>0.299</td>
<td>80</td>
<td>224.25</td>
</tr>
<tr>
<td>0.136</td>
<td>160</td>
<td>102</td>
</tr>
<tr>
<td>0.092</td>
<td>320</td>
<td>69.3</td>
</tr>
</tbody>
</table>

Source: Authors
4. Discussion

In many countries, brucellosis is a nationally notifiable disease and reportable to the local health authority but the underreported and official numbers constitute only a fraction of true incidence of the disease. Thus, the true incidence of human brucellosis is unknown and the estimated burden of the disease varies widely, from <0.03 to >160 per 100,000 population (Taleski et al., 2002; Pappas et al., 2006). It is estimated that the annual incidence of brucellosis in Saudi Arabia is 21.4/100,000 population (Pappas et al., 2006).

This study was conducted on 65 patients (42 males & 23 females) suspected of having brucellosis with Male: female ratio (2:1). All patients were presented at Prince Sultan Armed Forces Hospital at Al-Madinah Al-Munawrah, during the period of November 2015 to June 2016. The patients were examined using slide agglutination test for detection of anti-Brucella antibodies in their sera. The results illustrated in figure (2) revealed that 60 patients were brucellosis positive with a total incidence of 92.3%. Out of 42 male patients, 40 brucellosis cases were recovered with an incidence of 95.2% while 20 brucellosis cases were identified from 23 female patients with an incidence of 87%. On the other hand, the data presented in table (2) pointed out that brucellosis was seen in all age groups however its incidence varied. The incidence reached to 100% among the age groups (10-20) years old and (50-60) years old. These results generally agree with the previous reports of Mantur et al. (2006) who recorded that human brucellosis affects all age groups and males are affected more commonly than females which may be due to risk of occupational exposure. Also, these results agree with results of Hizel (2007) who concluded that brucellosis may be seen in any age group, but it still involves young and middle aged adults more frequently with predominance among males.

The investigation presented here showed a variation in the incidence of brucellosis from season to another. The most of brucellosis patients were recovered during the period from January to June with an incidence of 100% for all months except March was 83%. However, the examined patients during November and December were negative for brucellosis using slid agglutination test as shown in figure (3). Elbeltagy (2001) regarded the variation in the incidence of brucellosis among seasons to that a rainy season in an area would result in increased grass growth and thus influx of shepherds and flocks. However, Memish and Mahi (2001) recorded that the highest incidence of brucellosis in Saudi Arabia was seen in spring and summer seasons.

Many researchers discussed the transmission of brucellosis. They confirmed that the disease is transmitted from animals to humans by direct contact with such infected animals, consumption of their infected dairy products, or inhalation of aerosols (Kurdoglu et al., 2010). Other than the possibility of transmission through blood transfusion and organ or bone marrow transplantation, brucellosis is not readily transmitted between human beings (Khan et al., 2001). The occupational source of exposure predisposes the farmers, shepherds, butchers, laboratory workers, veterinarians and slaughterhouse workers to a greater risk of contracting the disease through inhalation of contaminated aerosols, contact with conjunctival mucosa, or entry of the bacteria through cuts and abrasions in the skin as a result of contact with infected animals or their products (Cutler et al., 2005). The non-occupational sources of exposure
include ingestion of infected meat, unpasteurized milk and milk products (Cutler et al., 2005). In recent years there has been a shift towards food borne brucellosis transmitted by raw animal product consumption in urban populations. For example, consumption of traditional Arabian/African delicacies such as raw liver can cause human infection (Malik, 1997). In this respect, we evaluated the effect of risk factors on brucellosis incidence. It can be concluded from figure (4) that there were different risk factors for brucellosis. Consumption of milk products, heating raw milk and milking animals were the most risk with an incidence of 100% followed by drinking raw milk with an incidence of 95%. However, cutting raw meat and animal contact were the less risk with an incidence of 80% and 67%, respectively. These results presented new data than Seimenis et al., (2006) suggested that animal contact still remains the predominant transmission mode in rural and nomadic populations, practicing traditional husbandry. The results of this study agree with the previous report of Al-Eissa et al. (1990) who studied 102 cases of children with proven brucellosis. He found that the main source of infection was the consumption of raw milk in 80% of the cases. On the other hand, inhalation of particles, accidental inoculation and ingestion or direct contact may be responsible for the potential hazard of Brucella to laboratory personnel (Charissis & Vassalos, 2009). A number of cases of laboratory acquired infections have been reported [23-27]. Laboratory-associated infections represent 2% of reported cases of brucellosis (Sewell, 1995; Fleming & Hunt, 2006; Collins, 1988) demonstrating the high risk of acquiring Brucella infection in clinical microbiology laboratories where these highly infective bacteria are handled. Also, Weinstein and Singh (2009) concluded that brucellosis continues to be the most frequently reported laboratory associated bacterial infection worldwide. Therefore, biosafety level (3) precautions are required to isolate Brucella in a laboratory (Charissis & Vassalos, 2009). Brucella seemed to be of prominent interest as it can be easily transmitted via aerosols to humans. The organism is believed to have a low infective dose in humans of only 10 bacteria and 50 to 80% of exposed persons will develop clinical disease. In this scope, Khan et al. (2001) reported that other than the possibility of transmission through blood transfusion and organ or bone marrow transplantation, brucellosis is not readily transmitted between human beings. Congenital brucellosis has been reported but it is rare. Recently, other routes of transmission have been identified such as infection through breast milk, having an infected household member (Sofian et al., 2008); through sexual transmission and the infection contracted by an obstetrician during the delivery of a transplacentally infected baby (Akçakuş et al., 2005). In the Kingdom of Saudi Arabia, camel’s milk may be sold on the main streets and is considered a delicacy. On the premise of increased freshness, the milk is provided frothy and warm from the camel. It is even much easier for campers in the desert to find providers of such kinds of camel or goat milk. In other parts of the Middle East, fresh goat’s cheese may be sold in supermarkets. If such cheeses were purchased and consumed without cooking, there would be a good chance of disease transmission (Memish & Balkhy, 2004).

Saudi Arabia is a vast reservoir of human brucellosis. Regional endemicity varies in Saudi Arabia, partly according to climate factors; a rainy season in an area would result in increased grass growth and thus influx of shepherds and flocks (Elbeltagy, 2001). The increased incidence of brucellosis in this country is also attributed to massive importations of
uncontrolled slaughter animals and inadequate quarantine procedures (Memish, 2001). The absence of health literacy, which could interrupt the direct link between animal and human disease, is also critical (Bilal et al., 1991). Brucellosis in Saudi Arabia is hyper endemic, with more than 8000 cases reported each year to public health authorities. During 1998, brucellosis ranked as the No. 1 reportable communicable disease (22.5%) in Saudi Arabian National Guard communities (Memish & Mah, 2001).

The annual cases of brucellosis in Saudi Arabia were 15,933 and 5781 in 1997 and 1998, respectively (Pappas et al., 2005). Few studies described the epidemiology of brucellosis in some parts of Saudi Arabia (Al-Sekait, 1993; Fallatah et al., 2005). The human brucellosis cases increased sharply during the period 1985-1990 from 4.9 to 69.5 per 100,000. The highest rate was recorded in 1988 (796 per 100,000). The infection was reported all over the kingdom, but with marked increase at Al-Jouf; Aser and Qasiem (Memish & Mah, 2001). In addition to that the incidence of the disease is high in the Central Najed region, around the city of Riyadh, an area where part of the population has a nomadic heritage which perpetuates the ingestion of fresh camel, goat and sheep milk (Al-Eissa, 1999; Hafez, 1986). Many measures to control the disease were implemented as early as 1983 in Saudi Arabia (Al-Ballaa, 1994). More recently, it is estimated that the annual incidence of brucellosis in Saudi Arabia is 21.4/100,000 population (Pappas et al., 2006). Morbidity in the Saudi population continues to be reported with increasing frequency from various regions of the country, particularly from the rural areas, and human infection is in the range of 1.6%-2.6 % (Alballa, 1995; Al-Nasser et al., 1991). Khan et al. (2001) reviewed 92 pregnant women who presented with acute brucellosis at a Saudi Arabian hospital. From 1983 to 1995, the cumulative incidence of pregnancy and brucellosis was 1.3 cases per 1000 delivered obstetrical discharges. The incidence of spontaneous abortion in the first and second trimesters was 43%, and the incidence of intrauterine fetal death in the third trimester was 2%. For all the above reasons and as human brucellosis has a serious medical impact in Saudi Arabia, the study was conducted to estimate the prevalence of brucellosis in a Saudi hospital in Al-Madinah Al-Munawarah and to determine the most prevalent species of Brucella (Brucellameliflentensis or Brucellabortus).

The diagnosis of brucellosis is based on a detailed history obtained from the patients and the isolation of Brucella spp. from blood cultures are accepted as the standard method. The definitive diagnosis of brucellosis requires isolation of the bacterium from blood, bone marrow or tissue samples. Although the sensitivity of blood culture method varies depending on the individual laboratory practices, quantity of bacteria in blood and the methods used, it changes between 15-70 %. Bone marrow culture is considered as the gold standard for the diagnosis of brucellosis, since the relatively high concentration of Brucella in reticulo-endothelial system enables the detection of the organism. In the present investigation, we tried to make blood cultures for the collected samples but unfortunately the Armed forces hospitals (Prince Sultan Armed Forces Hospital, Al-Madinah & Military Hospital, Riyadh) policy prevents Brucella culturing because it is highly infectious agent and the lack of biosafety cabinet level (3), which is necessary to do the culturing process. The standard tube agglutination test (SAT) is the most widely used serologic test for the confirmation of human brucellosis(Pabuccuoglu et al., 2011). We used rapid slide agglutination test for titration of
anti-Brucella antibodies of brucellosis cases. From data available in table (3), it is evident that anti-Brucella antibodies titers reach to 80 in fourteen cases, 160 in thirty four cases and 320 in twelve cases. Corbel (1972) said that the serum agglutination test remains the most popular diagnostic tool for brucellosis. Titers above 1:160 are considered diagnostic in conjunction with a compatible clinical presentation. However, in areas of endemic disease, using a titer of 1:320 as diagnostic may be more specific. On the other side, Acha and Szyfres (2003) reported that serum agglutination test (SAT) referred as the standard Brucella agglutination test is commonly used for diagnoses of acute brucellosis. However, 2-mercaptoethanol and complement fixation tests are used for chronic brucellosis, where active infection continues even though agglutination titers return to low levels.

Results of Brucella species detection using slide latex agglutination test are presented in figure (5). They illustrated that the most prevalent species among examined patients was B. melitensis (86%) while B. abortus was detected from only (6%) of examined cases. These results agree with the report of Elbeltagy (2001) who studied 137 brucellosis cases occurring in Tabuk Province, Saudi Arabia in 1997. Brucella agglutination titer of > or = 1/80, or rising titer plus history of typical signs and symptoms were considered evidence of infection. The incidence rate was 34/100,000, mean age 33.8 +/- 13.9 years (range: 3-72 years) and male: female ratio 1.8:1. The most common infecting agents were B. melitensis, B. abortus and B. suis.

Al-Dahkou et al. (2003) added to the previous drawbacks that serum agglutination tests have also a major drawback in that they are not suitable for patient follow-up, since titers can remain high for a prolonged period. However, Araj (1999) found that some of these shortcomings can be overcome by modifications such as the addition of EDTA, 2-mercaptoethanol, or antihuman globulin. Inaccurate serological results causing incorrect diagnoses are a continuous problem when testing for infectious disease agents in an outbred population of animals or in human beings. Because of the genetic diversity of populations, some animals will respond with low antibody levels to exposure to Brucella spp., resulting in false negative results. Other animals will respond with very high levels of antibody which may cause prozoning in some of the older assay types. High responders may also have elevated antibody levels to naturally occurring antibody caused by exposure to cross-reacting microorganisms. Exposure to cross-reacting microorganisms may also cause elevated antibody levels for various periods of time, some prolonged. Both scenarios will result in a false positive serological reaction, a major diagnostic problem in some areas where such microorganisms are endemic (Nielsen & Yu, 2010). For all these drawbacks, enzyme immunoassays (ELISA) which measure specific IgM, IgG and IgA antibodies, allow for a better interpretation of the clinical situation and overcome the false negativities/positivities which may arise in SAT (Alişkan, 2008).

Almuneef and Memish (2003) concluded that Enzyme-linked immunosorbent assay (ELISA) has been evaluated for many years for their diagnosis performance to detect serum antibody to brucellosis. Indirect ELISA typically uses cytoplasmic proteins as antigens. ELISA measures class M, G, and A immunoglobulins, which allows for a better interpretation of the clinical situation and overcomes some of the shortcomings of the serum agglutination test. In this respect, Rahman (2003) recommended using of ELISA for diagnosis of
brucellosis that it has several advantages when compared with other tests. Firstly, it has a
direct method of identification of specific antibody and therefore, it is not prone to false
positive reactions. Secondly, it is more sensitive than other the agglutination test and thus has
the potential to detect infected animals. Thirdly, the antibody enzyme conjugate employed is
able to detect all classes of antibody. A combine determination of all classes of antibody
allows accurate serological diagnosis at any stages of disease. Fourthly, ELISA results
provide an epidemiological tool for investigation the infective status of flocks. Also, Patra et
al. (2014) concluded that Serological tests are more sensitive as compared to blood culture.
ELISA is most sensitive and rapid for the diagnosis of brucellosis. A combination of IgM and
IgG ELISA was found to be most efficient for detecting brucellosis, among the techniques
evaluated. Therefore, in this study we estimated Brucella specific IgG and IgM antibody
levels in the sera of examined patients using ELISA. As shown in table (4 & 5) the sera of 38
(63%) brucellosis patients were containing anti- Brucella Ig G antibodies and 22 (37%) were
negative. However, the sera of 4 (80%) brucellosis free patients were containing anti-
Brucella Ig G antibodies and only one (20%) was negative. On the other hand, the sera of 50
(83%) brucellosis patients were containing anti- Brucella IgM antibodies and 10 (17%) of
them were negative. However, the sera of 2 (40%) brucellosis free patients were containing
anti- Brucella Ig M antibodies and 3 (60%) of them were negative.

The data presented in table (6) recorded 34 (57%) patients with brucellosis and only
one (20%) brucellosis free patients were containing IgG/IgM combined antibodies in their
sera. It is of interest to note that 6 (10%) patients with brucellosis were negative for both. The
aforementioned results threw more light on the appearance of Brucella specific IgM and IgG
antibodies in sera. Mantur et al. (2007) explained that during the first week of infection, IgM
antibodies against lipopolysaccharide antigens appear in the serum, followed by IgG
antibodies as early as the second week. Both antibody isotypes peak during the fourth week,
and the use of was associated with a decline of both IgM and IgG class antibodies. They also
recommended that Brucella specific IgM and IgG are still the most common and useful
measures for the laboratory diagnosis of brucellosis as they are faster and reduce risk of
laboratory acquired infections due to handling Brucella culture.

Recently, the awareness increased to the whole-blood interferon-gamma (IFN-γ) assay
as a quantitative in vitro assay for a direct read-out of Ag-specific cell-mediated immune
(CMI) responses to infectious diseases. Riber et al.(2011) confirmed that the IFN-γ assay is
robust in severe intracellular infections like Brucella. For this reason, we evaluated levels of
IFN-γ in sera of examined patients using ELISA to be used as a marker for brucellosis.
Interpretation of the readings is based on the tested serum sample was considered as positive
if its ELISA absorbance value exceeds the mean value of the negative control samples by
more than two standard deviations(Gast et al., 1997). By comparing ELISA absorbance
values of IFN-γ in sera of examined patients with the calculated cut off value (0.261) we
found that among patients with brucellosis, 9 cases were having positive levels of IFN-γ
while all brucellosis free patients were having negative levels of IFN-γ (table 7). The
sensitivity, specificity, negative predictive values and positive predictive values of ELISA
tests used in this study are shown in table (10). Finely, we compared between results of the
used serological methods in diagnosis of those patients with signs and symptoms of

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brucellosis. Results of SAT as well as ELISA (IgG, IgM and IFN-γ) were recorded and tabulated in table (8). 95%, 83%, 71% and 12% respectively were brucellosis positive males based on these tests. In addition to 87%, 74%, 52% and 17% respectively were brucellosis positive females based on these tests (figure 6). Also, examining of 40 male patients with brucellosis based on SAT using ELISA for the presence of anti-Brucella IgG and IgM antibodies and IFN-γ in their sera resulted in 85% were having positive IgM titers, 73% were having positive IgG titers and 12.5% were having positive IFN-γ levels. However, 80%, 45% and 20% of female patients with brucellosis were having positive levels of IgM, IgG and IFN-γ, respectively (table 9 and figure 7).

In addition to that, it can be concluded from tables (11) that there were differences in ELISA results between 1/80, 1/160 and 1/320 antibody titer groups. The highest (%) of patients having positive IgG and IgM levels in their sera were in 1/160 antibody titer group. It was also noticed that all positive IFN-γ levels were belonging to 1/80 antibody titer group. The mean IFN-γ levels (pg/ml) according to SAT antibody titers were 224.25, 102 and 69.3, respectively (table 12). Therefore, Araj (2010) found that in order to achieve the most reliable diagnosis of human brucellosis, it is recommended that a laboratory use a combination of two agglutination tests, namely SAT and indirect Coombs or SAT and Brucellacapt, or ELISA for IgG and IgM. This allows the detection of antibodies at different stages of the disease, since in the acute stage any test can be positive whereas in chronic, complicated or focal cases SAT can be negative while Coombs, Brucellacapt and ELISA IgG are positive.

5. Conclusion and Suggestion

In conclusion, in countries like the Kingdom of Saudi Arabia, where brucellosis is endemic; rapid, and sensitive, highly specific diagnostic methods are required to make early diagnosis and prevent resistance as there is an overlap in therapy. The gold standard method in this study was SAT.

Conflict of Interest

No conflict of interest

Funding:

This research work is funded by Prince Sultan Armed Forces Hospital at Al-Madinah Al-Munawrah, Saudi Arabia.

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